# Roots and their fungal partners

A chapter in: *Trees of the People*, by Alan R. Walker www.alanrwalker.com

A young seedling of birch grew from between small fragments of rock where a load of road-stone had been dumped beside a forestry road then neglected for several years. During that time a thin growth of seedlings appeared on the pile: birch and Scots pine, also New Zealand willowherb. The herb was a long way from its original home, but the trees were in a forest that had been flourishing naturally in northern Europe for hundreds of years. Little resembling soil could be found within the pile of recently quarried stone, just finer gravel, mineral grains and a trace of decaying vegetation. Yet the seedlings were healthy, with little competition from grass or moss, and well lit beside this wide road.



Seedling of birch, *Betula*, on stone pile, and at left an older seedling from same pile, showing roots and mycorrhizal colonisation.

I found it easy to lift out the birch seedling and shake off grains of gravel. Under a hand-lens appeared the fine roots branching off the main root, and numerous short root tips. Most of these tips bore bulging sheaths, each like the finger of a woollen glove. When I submerged the seedling in water, held in a white dish, fine contrasting threads became just visible as they spread away from the sheaths. The roots of the seedlings bore some companion organism, some symbiont, down there in that inhospitable ground. A species of fungus was firmly attached to the seedling's roots. Clearly the fungal sheaths and threads were of the mycorrhizal type – a fungus-root.



Roots of birch seedling as shown above. Most root tips are sheathed with mycorrhizas, as arrowed.

Spores from the mushrooms of a species typical of this sort of forest, possibly a poisonpie mushroom, Hebeloma crustiliniforme, had been shed into the air near the stone pile and here they developed and spread sufficiently to contact the roots of a potential symbiotic partner. Once there the fungal threads wrapped around the growing tips of roots and penetrated between the outer cells. Here the fungus started to extract simple sugars from the seedling, sugars coming from the photosynthetic activity of the seedling's few leaves. To establish and grow on that barren soil it seemed that the seedling must have gained something in exchange. Seeds of trees germinate where they land after being blown on the wind, washed away in floods, or carried by animals. If their parent population is to continue to flourish, the germinating seeds will need space for their leaves to spread into the sunlight and capture the energy of photons. Their roots will need to provide a system of tubes for water and nutrients to be sucked up to supply the leaves. A forest fire supplies both space and nutrients after it has devastated a stand of trees. Trees wind-thrown after a gale make space and disturbed ground, so within a year seedlings will thrive on the new light and more accessible soil. Then mycorrhizal fungi that grew with the old trees discover the new roots of seedlings to colonize as a source of their energy from the plant's sugars.

Even ground as bleakly barren as the slopes of a volcano, or rock and till dumped by a retreating glacier, becomes vegetated with tree seedlings along with lichens and mosses. Species of trees adapted to life on mountains or lands near the poles, dwarf birch (*Betula nana*) and alpine dwarf willow (*Salix reinii*) can establish a few plants that slowly mature. Soon enough threads of mycorrhizal fungi spread through the stony soil in search of nutrients and water and become symbiotic with these pioneer trees. Then any new seedling of the same tree species will encounter an established network of fungal threads. Rapid fungal colonization of these newly arriving seedlings boosts their growth and enables them to grow and spread faster than their parents.

### Root structure and growth.

A germinating seed of a tree soon uses up its own resources to grow a primary root, its radicle. The radicle rapidly penetrates the soil to start providing the shoot with water and nutrients. Then lateral roots emerge from the radicle and these in turn form branches, many of which spread close to the soil surface rather than penetrating deeply. At the tips of the roots a clump of cells, a meristem, continually produces new cells both behind and forwards. The lower new cells develop forward into a cap of loosely bound cells. These are fat with vesicles of a watery, gelatinous, material: a mucigel. As a growing root pushes through the soil it discards these cap cells, which disintegrate to form a blob of gel around the root tip, lubricating and easing its passage through the soil.

Behind a root's meristem new cells grow and differentiate into layers for separate tasks. Outermost is the epidermis, just one cell thick. From each epidermal cell projects a thin hair as a simple extension of the cell surface. Root-hairs are active at the tips of advancing roots, taking up water with its dissolved nutrients through this array of large surface area. As the root continues to branch, each meristem at the root tip provides cells for the bulk of the root, its cortex layer. Cells at the centre of the cortex specialize further into transport tubes (xylem) allowing water and nutrients to reach the shoot, and as a contra-flow down phloem tubes to conduct nourishing sap down from the highest leaves to the tips of roots. A root and its branches of a wind-thrown Douglas-fir, *Pseudotsuga menziesii*, showing how they divide finely and spread to penetrate soil.



Branching growth of the roots will never cease in the life of the tree. As the shoot grows the main roots need to radiate in search of nutrients and to support the shoot. But older roots also continue to produce branches close to the centre of the root system so that the soil here continues to be exploited. So spread of roots away from the shoot is limited, rarely more than twice the diameter of the mature leafy canopy. The roots branch, extend, do their work, die back and regrow within this perimeter. A researcher studying roots of a stand of seventy year old northern red oaks (Quercus rubra) in eastern USA examined samples of soil of one cubic centimetre. He found there one thousand root tips with a calculated total length of 2.5 metres. Although these fine roots occupied only three percent of the soil cube their surface area was about six square centimetres, without counting the huge surface area of all the root-hairs. Life in soil is physically harsh and the demands of the shoot are great, so new fine roots must be replaced rapidly. As the root tip advances it leaves behind a trail of spent mucilage and cells. Redundant root branches rot where they ceased to function. Under a typical forest in a temperate climate between one and five tonnes of dead roots are left each year in the soil of an area of one hectare.



The growing shoot of a young tree needs support against the force of gravity as it rises. Most of the water and nutrients required are found near ground level where rain and leafy detritus land. So most forest trees stand on a superficial base plate where they thrive on adequate nourishment but at the risk of wind-throw during every gale. Hundreds of trees may be toppled in a fierce storm, but for the reproductive success of any one mature tree the fate of being wind-thrown increases its chances of leaving successful offspring. Its own seedlings may establish and grow fast in the sunlit space and disturbed soils.

### Roots compared to leaves.

All rooted plants live strangely compared to other organisms: half up in the air, half down in the ground. This split life demands continual compromise from seedling to senescent tree. The branches of a tree expand freely into the air, constrained only by gravity, their supply of nutrients, and competition for sunlight by nearby trees. Air provides a uniform supply of carbon dioxide for photosynthesis and oxygen for respiration. Only the amount of water vapour in air varies significantly. Every individual tree in one area of forest is exposed to the same overall supply of the sun's photons during one season. A stand of trees of the same species receives similar supply of nutrients to their leaves. In this aerial environment the risks to leaves are herbivores large and small, blasting by wind, and abrasion from particles of ice. The biological characteristics of leaves, needle-leaf or broad-leaf, evergreen or seasonal-green, can be described in terms of well defined physical components of their environment. This ecological analysis of leaves has been termed the leaf economics spectrum (see 'Trees-Leaf-fall').



Root hairs on fine roots and root tips of birch seedling. LEFT shows root tip not colonized. RIGHT shows root tip colonized with mycorrhiza.

Roots conform poorly to any similar analysis. They are impeded by the solidity of soil. Within a mass of soil, such as one metre deep by one hectare wide, the nutrients may be fairly evenly distributed in a soil of wind blown dust such as loess. In soil made from mountains ground

down by glaciers and rivers there may be patches of volcanic origin, rich in mineral nutrients, whilst other patches are of sandstone made of grains that long ago were leached bare of nutrients. A small patch of soil may contain the rotting remains of an animal, a mouse to a moose. Here the bones and soft tissues are a concentrated source of phosphorus, calcium, nitrogen ... The list of essential nutrients for trees contains about fifteen elements: boron to zinc. The best that roots can do is grope through the soil at random in search of patches of high concentration of a wide variety of nutrient types. A constant expense of carbon and nutrients to the ever growing and dying roots must be borne by tree's leaves. At least roots are protected from the browsing of large herbivores and massive infestations of insects, but soil contains small mites, millipedes, and similar animals that eat fine roots, and there are wood-rotting fungi that roots need to defend against. So researchers aiming to analyse roots in terms similar to leaf economics spectrum have instead demonstrated how different is the economic ecology of roots compared to leafy branches. Then there are the symbiotic mycorrhizal fungi to consider.



Constraints on roots compared to leaves.

## Soil and fungi.

A volume of fertile soil typically has about forty five percent as inorganic grains of rock. Five percent is the organic component: live roots, fungi, worms, mites and insects, protozoa and bacteria, all mixed in with the decaying remains of these organisms, plus leaf litter and stems of trees, herbs and grasses. The remaining half of the volume comprises air spaces and water. Except for roots and earthworms, most of this enormously varied living component of the soil is unseen by silviculturist or gardener, unless they come equipped with core-borers, sieves, extractors, and a microscope. There is however one of the living components of soil that regularly makes itself conspicuous: mushrooms. These are the reproductive bodies of fungi, their fruiting bodies or sporocarps. These create and disperse potential offspring as minute spores. For every sporocarp rising above-ground there spreads a network of fungal threads in all directions of the upper layers of soil. Here most nutrients can be found.

Oyster mushrooms, *Pleurotus* ostreatus. The sporocarps of a fungus that feeds on wood, usually as a saprotroph on stumps of dead trees but may also feed as a parasite on live trees. Credit: Wikimedia, H. Krisp.



Woodlands and forests are home to many thousands of species of fungi. None of these fungi possess chlorophyll and no fungus ever evolved into a symbiosis with the cyanobacteria that became the chloroplasts of all green plants (see 'Photosynthesis'). So fungi cannot directly obtain energy from sunlight. Instead, early in the history of life, they evolved a suite of powerful enzymes that digest all manner of the most abundant food: dead organisms of every kind. Most fungi still feed as *saprotrophs* (from sapro- for decaying matter; also called saprophytes, saprobes or saprophages). Other species of fungi can attack and feed on live cells and organisms. These are *parasites*, their effect on their hosts is pathogenic, causing harm that is often defined as a particular disease of plants or animals. A typical saprotrophic fungus is the oyster mushroom, *Pleurotus ostreatus*, which naturally feeds on dead trees and cut timber. However, the enzymes that the fungal threads secrete onto potential food are powerful enough to enable this fungus sometimes to feed on living trees, as a parasite rather than a saprotroph.



Spores of *Pleurotus ostreatus,* oyster mushroom. Scale bar is 10 micrometres; fungal spores are very small. Credit: Wikimedia, Alan Rockefeller.

Under most stands of trees there will be scores of species of fungi in mycorrhizal symbiosis with the trees, herbs and grasses. The sporocarps of some of these species, as they emerge at the end of the plant's growing season, are a splendid sight. Amongst the best is the fly agaric Amanita muscaria, typically under stands of birch trees, Betula species. Although the combination of live plant and fungal material and their decaying remains in the soil is small in terms of the volume of soil, their biomass in the soil as kilograms per hectare of forest ground can be a high proportion of the total biomass within a forest, above and below the ground. A study of stands of Pacific silver fir, Abies amabilis, found that of this total biomass (non-animal) eighty eight percent was in the soil. Although only six percent of the below-ground biomass was living and just a small fraction of the live matter was identified as mycorrhizal fungi, these fungi were responsible for fifteen percent of the combined net primary production of the assemblage of trees plus fungi. Most of the carbon that supported this growth was from the photosynthate of the fir trees. A significant proportion of the carbon captured by photosynthesis by needleleaves went to support respiration and growth of fungi.

There is a common type of mycorrhizal fungus described as *arbuscular* (from a tree-like part of their structure). They are in the large group Ascomycota (a phylum or division, depending on which taxonomist you ask). Fungi are classified as an entire Kingdom of life forms. (Other Kingdoms are: Plants, Animals, Bacteria, plus another three of lesser known organisms.) Fungi are only plants in popular speech – technically they are no more plants than are animals or bacteria. The arbuscular mycorrhizal fungi are symbiotic with many types of plant. Their fine spreading threads, their hyphae, advance through the soil, contact the roots of a suitable plant then squeeze through the spaces between cells of the epidermis and cortex. The hyphae also penetrate through the cellulosic outer wall of the plant's cells and out the other side. They achieve this without piercing the plasma membrane of the plant cell. Instead the plasma membrane forms an invagination, it folds in on itself, so these violated cells remain alive and with functional cytoplasm and nucleus.



Simplified representation of an arbuscular mycorrhiza colonizing a root. Hypha (grey) penetrates an epidermal cell then grows through or around cells of the cortex, and finally forms an arbuscule inside a cortex cell.

Once in a suitable cortex cell the tip of a hypha will branch and curl into a tangled mass. This develops within the boundary of the cell whilst remaining outside the living plasma membrane of the cell. This structure, the arbuscle, sits in intimate contact with the metabolic apparatus of the cell, absorbing the plant's sugars. After four or five days the arbuscle withers and the colonized cell slowly recovers its normal appearance and function Meanwhile nearby root cells are being newly colonized. Another group of mycorrhizal fungi belong to the group called Basidiomycota. They commonly live in woods and forests of temperate climates in north and south hemispheres. These fungi are the ones that wrap around as tight sheaths on the root tips of many species of trees (photo page 2). Hyphal threads push down between the outer cells of the root, but do not penetrate the cells. These are the *ectomycorrhizas*, always external to the cells of their plant partner.

Representation of a root tip in cross section showing colonization by an ectomycorrhizal fungus.

The root is of a broad-leaf tree. The fungal threads from the mycelium penetrate between the epidermal cells to form a Hartig net, arrowed. These fungal hyphae do not penetrate inside epidermal or cortical cells.



Mycorrhizas are an ancient form of life, researchers have found them as fossils in the hard rock known as chert, a material our ancestors made into hand-axes and spear points. Chert from several sites, Scotland to Antarctica, contained the characteristic arbuscles of ascomycete fungi. These fossils were dated as formed in the Devonian period, four hundred million years ago when the land was becoming well vegetated with photosynthesizing plants for the first time. The earliest of these plants, types of moss for example, had no roots and possibly this was when an association with fungi evolved that developed into a symbiosis. Fossils of basidiomycete mycorrhizal fungi occur in the same type of rock, found in British Columbia but dated as from only fifty million years ago during the Eocene epoch. During those times there were forest trees, needle-leaf and broad-leaf, covering all lands of Earth except the coldest and driest in those generally warm and wet times. These fossils show the tips of roots, probably belonging to needle-leaf trees, covered in mats of fungi.



Hyphae of ectomycorrhizal fungus spreading as a mycelium around fine roots of birch tree.

Root tip arrowed has a sheath of the fungus actively colonizing it.

## Ectomycorrhizas.

Ectomycorrhizal fungi become familiar once their connection with characteristic woodland mushrooms is known and this type will remain the focus of this chapter, for simplicity and because they are the commonest type of mycorrhizas on forest trees in much of the world. A hypha is a single thread, and masses of hyphae spread through soil as a network called a mycelium. Hyphae construct their outer walls mainly of chitin, combined with other materials including cellulose. Chitin is the same substance that animals such as insects use for their external skeleton – tough and flexible. Hyphae may form with walls of separate cells or they may form into exceedingly fine and long open tubes. Within the hyphae are always true nuclei, membrane bound structures of the type called eukaryotic. This is one reason why the many species of single-celled fungi are distinguished from the bacteria. Bacteria always have a type of nucleus that is diffusely spread within the cell, as a prokaryote.



Hyphae of a fungus spreading across soil. These fine structures make up the main mass of many species of fungi that live in soil. From such hyphae the fruiting bodies, sporocarps, develop as the familiar mushrooms that will produce reproductive spores. Credit: Wikimedia.

Hyphae may be as narrow as 0.5 micrometres diameter, visible only under a microscope at high magnification, or as wide as 100 micrometres, which is twice the thickness of a human hair. Hyphae grow rapidly from their origin as a single spore, lengthening from their tips as they spread in all directions. The mycelium that develops is a genetically identical organism, a genet, either an individual or a clone. This organism may grow and live for decade after decade, sometimes growing into a thing weighed by the tonne. The famous humongous fungus, not the original specimen but the one currently claimed as the world record holder for size, is a honey fungus, *Armillaria ostoyae*. This species is familiar from its ordinary brown sporocarps clustered at the base of conifer and broadleaf tree stems. There it is parasitic, not mycorrhizal. This specimen, in the USA, is estimated to cover 880 hectares of forest floor, to weigh 600 tonnes and to have taken 2400 years to grow from one minute spore. Who knows, are there bigger fungi down in the woods to discover?

Ectomycorrhizal fungi reproduce sexually, using a genetically varied and complex method to produce spores for dispersal. These individual fungi

need to disperse their offspring far away from their mycelial network in the soil. Offspring need fresh soils to forage in, new plants to colonize and new fungi to reproduce with. Mycelial hyphae bind together to construct a sporocarp as a stiff body that pushes out into open air. Spores are produced by the trillion from various sporocarp structures. The ectomycorrhizal Amanita muscaria is typical, with the underside of its mushroom cap grown into a dense array of thin white gills that hang vertically. The dark gills of the commercial edible mushroom sold by greengrocers, Agaricus bisporus, are easily seen. At the surface of gills the tips of hyphae protrude as sexually maturing basidia. These bud off repeatedly as oval spores, smaller than the diameter of a human red blood cell. On a calm day many spores will land on soil occupied by their parent fungus, to no effect, but a light wind will carry millions of spores up to several kilometres away. Here there may be a gap of disturbed ground with few fungal competitors and some seedling trees to colonize. You may have seen fungal spores like dark smoke when pumped out of a mature puffball sporocarp (Lycoperdon species and others) as heavy raindrops land, or by gently poking a mature puffball that has its opening on top.



Mature sporocarp of *Amanita muscaria*, the fly agaric mushroom, showing the spore forming gills on the underside of the cap. These fruiting bodies of the mycorrhiza developed directly from the underground mycelial network.

### Symbiosis: the losses and gains of living together.

Mycologists have found and named one hundred thousand species of fungi and reckon there are as many still waiting discovery. At one thousand new species formally described in journals of taxonomy each year, and the undiscovered ones being more cryptic, that could take a while. Fungi are a predominant life form because their basic method of obtaining food is to rot down dead organic material. There is much of this stuff: leaves, roots, stems, corpses of animals from earthworms to deer. This rotting by fungi, this saprotrophy, must have been earliest method of feeding to evolve in fungi based on the concept that to become a symbiotic fungus it is necessary to start as a fully independent organism of a particular species. Then a partnership with some other organism may evolve, such as fungi living together with algae as the symbiotic life-form called lichen. Both partner species must change greatly. The mycorrhizal fungi of both arbuscular and ectomycorrhizal types have been coevolving with their tree partners for sufficient millions of years to have lost some of their ability to feed as saprotrophs. If they have lost most saprotrophic ability they need a plant as a symbiotic partner, thus they are in obligatory (or obligate) symbiosis. If they can also feed as saprotrophs they are in optional symbiosis, technically known as the facultative type, and can manage without a plant partner if necessary.

This becomes more complicated but more interesting. A *symbiosis* is between separate species reproducing independently whilst living close together in some functional relationship. Symbiosis refers to the physical location of the species, without implying gain or loss. With the type of mycorrhizal fungi and trees that are the focus of this chapter, their relationship with each other is fairly specific: one species of fungus may partner up with several species of tree, similarly for one species of tree and other species of fungal partners. But always both partners have evolved modifications of form and physiology that enable their symbiosis to work. These paired sets of genetically determined modifications vary along a spectrum of benefit to one partner relative to benefit to the other partner. If all the benefit goes to one partner then that partner is a *parasite* and the other partner in this symbiosis is its *host*.



Representation of two root tips and an ectomycorrhizal mycelium. Left: root tip with functioning absorptive hairs and not colonized. Right: root tip colonized and mainly absorbs nutrients from the fungal mantle.

All parasites are symbiotic by definition – a louse lives on the skin of its bird or mammal host, feeding on blood. So the term symbiosis has nothing to say about sharing of resources – the term is neutral about the economics of costs and benefits. If the harm to the host partner can be measured and defined by silviculturists then a disease is caused and is given a name, such as ash die-back caused by the ascomycete fungus *Hymenoscyphus fraxineus*. In the case of mycorrhizas and trees it would be the tree that is harmed because the fungus has colonized the tree. When both partners of these symbioses gain benefits from each other that are in a dynamic balance, with approximately equal benefits to both partners, the symbiosis is a *mutualism*.

Mutualism usually develops between two types of substantially different organisms that, because of their separate evolutionary histories, have

attributes that the other partner lacks and needs. Mutualisms are not necessarily symbiotic as in living closely together. Species of flowering plants often have evolved a mutualism between animals that aid cross pollination of the plant. Some of these relationships are intimate, such as species of orchids with flowers that mimic the shape and colour of particular species of bee. The bee attempts to mate with the orchid which responds by covering the bee with pollen. The frustrated bee then searches for another potential mate. Many flowering plants have a looser mutualism with a variety of mobile insects, birds and mammals. These different species of pollinators form a mutualistic *guild* of animals all with a taste for sugary nectar. Many flowering plants have evolved to provide nectar, easily available at the base of the flower to attract any member of a guild: bee, beetle or bird. Animals of various species will individually push past an array of pollen-bearing anthers of the single species of flowering plant to reach its nectaries.

Mutualisms arise through long, slow, coevolution of interacting adaptations by natural selection acting on two species. Where there is a close symbiosis, as with mycorrhizal interactions with the structure of plant roots, a single species of fungus has separate mutualisms with various species of plants. As with the example of pollination, this mutualism may have coevolved with a guild of plant species. In the case of pollination the resource flowering plants have to offer members of their animal guild is carbohydrate. In the case of mycorrhizal fungi the resource they have to offer to members of their guild is mineral nutrients. Thus a mycelial network of threads of a genetically single fungus connects to individual plants of several species as a *common mycelial network*. This can be an important component of how a wood or forest thrives.

Examples of direct and close three-way symbiotic mutualisms between three separate species are not evident. Do they exist? This question relates to the expanding complexity of the necessary channels of information for evolutionary selection: 2 partners have 2 channels; 3 partners 6 channels; 4 partners 12 channels . . . The photosynthetic cells of plants contain two symbiotic organisms that are in mutualism with the plant cell: the minute organelles called chloroplasts and mitochondria. But how far did these two formerly free-living microbes coevolve with each other when in the plant cell? (see 'Photosynthesis'.)



Representation of a tree seedling colonized by an ectomycorrhizal fungus, all within a laboratory microcosm. Roots (yellow) spread and forage for nutrients. Fungal mycelium (white) spreads from a site of inoculation and forms mantles around many of the root tips.

If both partners gain a separate benefit from their symbiotic partner then both partners are mutualists. But what if one partner gains more benefit than the other, what if the ratio of cost to benefit is not 1:1? Does the cost of producing sufficient sugars by photosynthesis in the tree to provide these large masses of fungi with structural and energetic materials have to be balanced equally in some way with resources from the fungus?

Nutrients for plants are usually in short supply. If the plant can extract nutrients from the fungus without harming the fungus then the balance of mutual benefit may be maintained. But how to compare sugars that travel from tree to fungus with nutrients such as nitrogen that travel from fungus to tree? The economic value of a tree's sugars to the fungus can be measured directly in units of energy available (as joules, or as kilowatt hours), or units of mass of construction carbon (grams). The efficiency of use of carbon by the mycorrhizal plant can be calculated by dividing the mass of carbon gained by the plant by the mass of carbon provided by the plant as sugars to the mycorrhizas. Nitrogen however is an element that enables respiration and construction indirectly. Similarly for other crucial nutrients such as phosphorus that come in extra amounts to the plant through the mycorrhizas. The problem is how to relate metabolic effects of nitrogen and phosphorus to gains of carbon.



Seedling of Scots pine (*Pinus sylvestris*) from same stony site as the birch seedling shown in page 1. Its roots were colonized by an ectomycorrhizal fungus which might have colonized seedlings of these two tree species.

The criterion for balance of benefits that works from an evolutionary perspective is the fitness of both partners. This biological fitness is defined as the ability of a particular genetic or physical type (genotype or phenotype) of organism to produce more offspring relative to other organisms of the same species. Natural selection will probably then lead to more of that new type of organism. The variants arise from random mutations that change the information transmitted from parent to offspring by DNA. The mutations arise either spontaneously within the organism or are induced by external influences such as natural radiation. Those variants most likely to prosper are an infinitesimally small subset of all possible variants, but because the mutations arise randomly and frequently there is within nature a constant bubbling up of slightly new designs of potentially fitter organisms – fit in this evolutionary sense. Living things flourish through their drive to reproduce themselves, by being adaptable in how they survive and reproduce.

Researchers have studied growth rates in many combinations of tree and mycorrhizal fungus, grown together in simple apparatus in a laboratory. They construct a chamber, a microcosm, from a pair of small transparent plastic plates fixed five millimetres apart. Bigger kit of this sort is called rhizotron. After filling the chamber with manufactured soil and planting a seedling, and either inoculation with mycorrhizal fungus, or without the fungus as negative control, then seedling growth is compared. Growth rate as gain of seedling weight during a fixed time can be measured for many combinations of tree and fungus species. By comparison of many studies (meta-analysis) researchers showed that in forty eight percent of such experiments there was a positive effect of mycorrhiza on seedling growth, whilst twenty seven percent showed no effect and twenty five percent showed a negative effect.

Were these negative effects caused by mycorrhizas on those seedlings behaving as parasites? Possibly they did, but to prove that researchers would need to expand their studies to measure both fitness of the trees and fitness of the fungi. For the trees these seedlings would need to be nurtured to reproductive maturity at twenty years or more, whilst the control trees would need to be kept free of mycorrhizas for that time. Only then could fitness be compared by testing viability and vigour of the new seeds and trees. We are not likely to have an answer any time soon. Never mind: this analysis shows that the concept of mycorrhizal symbiosis may be most productive if it includes the potential for both parasitic and saprotrophic behaviour by the fungi. Life thrives by its endless spontaneous variation in ways of reproducing itself – completely indifferent to how we humans try to define and categorize it.



Three sporocarps of fly agaric at different stages of maturation in a stand of birch trees: a mycorrhizal symbiosis easy to see at this level.

Saplings from a nursery had been planted on abandoned pasture for livestock. The nearest mature birch wood is 1km distant. These trees may have been first colonized by mycorrhiza from the seedling nursery, or colonized by windblown spores from sporocarps in the distant birch wood.

### Foraging for nutrients.

Viewed under a stereo-microscope a sample of a tree's roots will probably show a few root tips freshly pale, bearing root-hairs. The remaining root tips will be covered with a dark dense matt of hyphae, as a sheathing mantle. Spreading from the mantle will be hyphae and further from the mantle are hyphae from other mantles that combine as a mycelium. Nothing of the root tip can be seen: the fungus dominates by its colonization. On a flowering tree, an angiosperm, the hyphae squeeze between the cells of the epidermis and no further, but on a coniferous tree, a gymnosperm, the hyphae push in amongst the cells of the root's cortex. In both cases the resulting mesh of hyphae is named the Hartig net. No evidence of defensive modifications by the tree roots can be seen, no thickenings or suberization of the epidermis or cortex cells. For any one symbiosis of fungus species and plant species this is a dynamic and variable relationship. Ordinary non-colonized root tips emerge, grow, absorb nutrients and water, all in few weeks then they die. For colonized roots the same tempo proceeds, three to ten weeks is typical for the life of a mycorrhizal mantle. From the mantles the mycelial hyphae grow at two to three millimetres daily. This is visible when seedlings are grown in microcosms. Hyphae intertwine and thicken into strands, braided as a white rope. Under a forest floor these hyphal strands, growing like roots, are called rhizomorphs. They radiate out metre after metre, with fans of separate fine hyphae at their periphery, foraging at random for nutrients and water. Once a good patch of resources is found the hyphae concentrate around it densely. Roots continually grow from their fine tips, become colonized and engage in short-lived mutualist exchanges, to be replaced with more root tips that continue to seek nutrients and water. Growth, movement, temporary exchanges, senescence, death and replacement by fresh growth is the life of roots and mycorrhizas - a dynamism hidden under the soil surface.

Estimates of the mass of mycelium in the soil under a forest vary from the high hundreds to several thousands of kilograms per hectare. To that must be added the seasonal crop of sporocarps above-ground. Typical data from one study, in a Swedish forest of Scots pine (Pinus sylvestris) and Norway spruce (Picea abies) colonized with ectomycorrhizal fungi, showed a mycelium turnover of 125 to 200 kilograms of per hectare per year, and similarly 6 kilograms of sporocarps. The total biomass of these mycelia in the soil was 600 to 900 kilograms per hectare and this estimate did not include the mass of fungal mantles on roots or aboveground sporocarps. These estimates of biomass of mycelium are in the same range as estimates of the biomass of roots. Walking through a forest we find it difficult to comprehend these comparisons of biomass of trees and their partnered fungi - too counter intuitive. We people of the woods, where sunlight shines through green leaves and dapples the ground, can only dimly imagine life from the perspective of a mole burrowing in search of worms.

Of the total respiration of the symbiotic root system of a colonized tree thirty percent can occur in the fungal partner. All of the energy used by the fungus comes from the photons captured by leaves. Energy flows from leaves as sugary sap down the fluid transport system of phloem tubes and out to the root's cortex and epidermal cells. There the sucrose in the sap diffuses to the fungal hyphae. The plant is the source of the sucrose whilst the fungus is the sink. Within the outer membrane of hyphae are embedded arrays of transporter molecules (sucrose transporters, SUTs, and monosaccharide transporters, MSTs) that actively shift and transform this plant food for use within the mycelium. The enzyme fructase within the hyphae splits the sucrose into fructose and the glucose that the fungus uses for respiration and growth.



Remains of a red deer in a pine forest: a concentrated pool of nutrients will form in the soil below.

The benefit to the trees from these below-ground fungi comes as nutrient minerals, borne in watery solution as ionic compounds such as ammonium ion bearing nitrogen, and phosphate ion bearing phosphorus. There are about thirteen other elemental nutrients required but in smaller amounts and some may be easily available. These forms of nitrogen and phosphorus are naturally absorbed through the root-hairs but are frequently in short supply to the tree because they are scarce in the soil, or the roots cannot forage sufficiently to meet the demands of shoot and leaves. Nitrogen is the most important for plants by quantity, essential to construct amino acids, proteins and nucleic acids. Ammonium ion is usually the major source of nitrogen in forests. The mycorrhiza *Paxillus involutus*, on conifer and birch trees, contains ammonium ion in its mycelium. Trees in symbiosis with *P.involutus*, being short of nitrogen, act as a sink of this resource, drawing it in by diffusion and differences in concentration gradients across cell membranes. Alternatively, nitrogen is actively transferred across cell membranes of cortex and epidermal cells by specific transporter molecules (ammonium transporters, AMTs).

If a nutrient such as nitrogen or phosphorus is scarce then the plant may have a relative excess of carbon available to its mycorrhizal partner. In contrast, if the plant happens to be on soil rich in the nutrient element then its potential for growth may be diminished by loss of too much of its carbon to the mycorrhiza. The precise ecological context of both soil and above ground climate conditions can strongly influence the balance between loss and gain in this symbiosis. Phosphorus is often in severely short supply for plants. Yet to grow and thrive plants need phosphorus to construct the phospholipid molecules in the membranes of all cells, for the sugar-phosphate molecules such as adenosine triphosphate for respiration, also to construct components of nucleic acids. The hyphae of ectomycorrhizal fungi secrete phosphatase enzymes that digest organic debris and absorb soluble phosphate ions. The tree's roots absorb the phosphate in ways similar to that of nitrogen. This occurs at the other end of the mycelial network from metres away where concentrated patches of nutrient may occur. Phosphorus in ionic form is actively carried across cell membranes of cortex cells by specific transporter molecules (inorganic phosphorus transporters, Phts).

Researchers using a radioactive isotope of phosphorus, <sup>32</sup>P, can trace the movement of phosphate within individual hyphae from a concentrated spot in artificial soil all the way into a tree seedling. Radiation from the isotope reacts with photographic film, revealing as white streaks in the mycelia and roots of laboratory microcosms. Phosphorus concentrates around the roots of *Pinus* seedlings colonized by *Suillus bovinus* mycorrhiza. These experiments show an increase in transfer of phosphorus

from soil to colonized plant at x25 to x40 the level of control seedlings without mycorrhizas. Ectomycorrhizal fungi may also have similar abilities to those of lichens that grow on rocks. There is evidence that mycorrhizal hyphae secrete substances such as oxalic acid that etch the surface of rock such as apatite, containing compounds with phosphorus. The significance is uncertain for such active weathering of rock by mycorrhizas, in comparison with their known ability to extract more soluble sources of phosphorus in the soil under forests.

### Forestry and mycorrhizas.

The term 'Mycorhiza' was invented by the German botanist Albert Frank in 1885, but now this field of research has been modernized by application of molecular genetic techniques. These enable researchers to follow fungal activity in soil as it happens in the forest in addition to what they can observe in the laboratory. Molecular markers have been developed for many uses in biology (complementary DNA microarray; gene sequencing or profiling; polymerase chain reaction; analysis of microsatellite DNA loci . . . ). These can be adapted in the lab for use on known species of fungus in pure isolation, or in culture, or from their sporocarps. Now researchers can plot the distribution and activity of an individual fungus that grew from a spore. This genetically defined organism, or clone, is called a genet. The trees the fungus is symbiotic with are surveyed as they grow in a forest stand. With sufficient number of samples from soil-borers, researchers make maps of the natural distribution patterns of mycelia of different fungal genets.

One study of a stand of sixty seven Douglas-firs, *Pseudotsuga menziesii*, in south west Canada was sampled intensively for mycelia containing the genets of thirteen individuals of the mycorrhiza *Rhizopogon vesiculosus* and fourteen individuals of *R.vinicolor*. These species develop rhizomorphs growing tens of metres through forest soil. Their individual mycelia formed symbioses with many individual trees. The complex interconnections created a common mycelial network. The greater the age of any single tree the better connected it was, often to trees many metres away.

# Roots and their fungal partners



Simplified representation of a common mycelial network of hyphae and rhizomorphs that inter-connects trees of two species in a forest. Fungi are all of one species but of two genetically different populations (genets) shown as red and blue inter-connections.

Of the 33 trees shown the large conifer at bottom left is the dominant hub tree of this group. (Based on: Beiler, 2010.)

Recently three researchers on mycorrhizas published a paper in a leading journal, strongly criticizing the content and conclusions of a group of research publications about how mycorrhizas and trees in a forest interact (Karst 2023, see References and notes). This critique focused on common mycelial networks, where the mycelia interact with more than one plant, as described in this chapter on page 17. Karst and co-authors focused on three claims made for these types of network: that they are widespread in forests; that nutrients are transferred within the network such that survival of seedlings is enhanced; that mature trees selectively provide seedlings resulting from their own reproduction (offspring or kin) with nutrients and defence signals via the network. These authors found insufficient evidence from field and laboratory studies to support these interpretations of the activity of mycorrhizal networks as a widespread support network between trees. To me this controversy shows a difference of approach of researchers about how to understand interactions of the many species living in a definable area of land or water. A wood or a lake for example, is obvious to us as a separate physical thing. This is a problem of complexity in science. The domain of live organisms is in constant activity as each individual responds to its vital drive to reproduce itself. The more species of organisms are in one place the more complex that group becomes. To get a grip on this some researchers call a wood or a forest an ecosystem, with stress on systems and patterns of collective activity that can be detected. Other researchers call a wood or a lake a *community*, or an assemblage. These three terms are human concepts, products of our thoughts. Ecosystem assumes that the whole is greater than the sum of its parts because it works together in a systematic way. Community suggests there are some systematic components within the forest, whilst assemblage is deliberately non-commital about system and focusses on how individuals and species grow and reproduce. Also the term ecosystem is used as a loose catch-all to describe a group of organisms by where they live rather than how they work together. Furthermore a newer concept, complex adaptive system, is being used in the same way as ecosystem is meant in its strict sense.

Supporters of the ecosystem concept advocate a holistic approach based on discovering connections and linkages between many components of a forest: trees through to the animals that live in soil. Supporters of the assemblage concept use a reductionist approach in which the interactions between a few species are studied at the level of individual organisms and with emphasis on the aspects of their evolutionary fitness for reproduction. The holistic approach leads towards analysis of varied data involving many linkages and often requiring mathematical models to gain understanding. The reductionist approach leads to hypotheses testable by experiments or simple observations, and routine statistics, on several species at a time. Reductionist methods tend toward failure to see the forest for the trees. Which is best? How can researchers study forests with the aim of maintaining the health and continuance of natural forests, or their productivity as managed forests? In this book I use the approach typified by the term assemblage, as used commonly in research papers and explicitly in textbooks on ecology such as Krebs 2014, and Colinvaux 1993. My aim for this book is to report on recent results of researchers whilst remaining grounded on textbook knowledge. For now, this topic is too broad and controversial to include here. Despite these conflicts both these ways of thinking are useful for forestry practice. So anther chapter of this book, will return to this topic in the context of deforestation in transition to reforestation.

All the trees in a stand are potential sinks for scarce nutrients that the mycelia contain. The faster the tree is growing the stronger is its relative strength as a sink toward which nutrients travel. Seedlings of these trees have a fundamental imperative to grow fast to establish themselves in competition for light down there at ground level and in competition with herbs and grasses that need the same space and nutrients. The more rapidly and robustly a seedling can gain its own symbiosis with this common mycelial network the greater its chances of survival and ultimately its reproductive success. Many species of ectomycorrhizal fungi are fairly flexible about what species of tree they partner with. Where there is open ground that both mycorrhizal spores and tree seeds may by chance land, this low symbiotic specificity is an adaptive advantage to both partners. The probability of rapid establishment of mutualism is increased. The higher the local diversity of species of fungi and of trees the better it will be for the benefits of mycorrhizas.

This increasing knowledge of mycorrhizas is being used in various ways for forest management and silviculture. The commercial value of the largest and straightest tree trunks in an area for harvesting is obvious. Felling such trees (called high-grading) in preference to lesser trees during harvest of forest will remove the ecologically most important trees, the hubs of the common mycelial networks. So for the long-term productivity of wide areas of forest sufficient hub trees should be left as a source of mycorrhizal fungi into replanted areas nearby. Mycelia remaining in the soil after clear felling survive for no more that several years without their tree partners. They do not fruit as mushrooms without their trees and their saprotrophic capacities are insufficient to support continued independent growth or reproduction. To maintain these mycelial networks refuge areas for trees can be left in harvested areas, life-boats for plants and fungi. A combination of hub trees, as various species of both needle-leaf and broad-leaf trees, should be maintained in an area sufficiently large to be a self supporting community. The shape of the refuge will be best if it is complex, with a high ratio of edge to total area, a thin strip rather than a square. This will increase the spread of mycelia into harvested areas over a wide front.

New plantations should contain a range of species of conifers and broadleaves so that a wide diversity of mycorrhizal species can be supported. One study that compared forest stands of different ages found that sixty five years were needed for full fungal diversity to re-establish after widespread clear felling. Useful fungi deserve management as long-term as do the trees. For replanting with seedlings this should be done as early as feasible in the felled areas, within one or two years to gain from the existing mycelial networks surviving there.



Granules of mycorrhizal treatment for application to soil. Granules are approx. 3mm across and consist of fungal hyphae and spores in an inert carrier. This is a small packet sold for gardeners; similar formulations are manufactured and sold in 20kg bags or larger for commercial use. Additionally the seedlings can be cultivated in nurseries where mycorrhizas are introduced as inoculants. Some fungal species are grown in artificial media on industrial scale for use in horticulture and tree nurseries. The fungus Thelophora terrestris forms symbioses with species of pine, spruce and oak when in nurseries. Inocula of Pisolithus tinctorius, Rhizopogon vinicolor, R.colossus, Hebeloma crustuliniforme, Laccaria bicolor and others are also used. With these ranges of fungi suitable for cultivation the best effect will be where the species of fungus matches what is usually found with the plantation trees. However, the relatively low specificity of some mycorrhizas for their guild of partner trees provides useful flexibility. From the liquid of fungal cultivation media masses of hyphal fragments and spores are dried out and mixed with an inert carrier material such as fine sand or vermiculite. In this form manufactured and packaged inoculant can be spread broadly over the soil. Many commercial companies supply these products, often packaged in large bags for forestry and horticultural use.

Another silvicultural trend effective for maintaining thriving populations of mycorrhizal fungi is called continuous cover forestry. This is specially relevant for deliberate restructuring of plantation forests so that they develop with a varied age structure and with few or no large areas to be clear-cut. This is most relevant for countries where there are few or no remaining forests for timber production that are derived directly, by forestry management going back to when the forests were natural. More will be written in this book in further chaptes in preparation.

Symbiosis is a topic full of mysterious connections and the complexities of studying theories of coevolution. As if office-bound theory was not problem enough, out in the forests these diffuse, protean, dynamic, tiny and vastly huge organisms live underground with their tree root partners in soil all dark, dirty and teeming with other life competing for nutrients, water and space. Despite all these impediments mycorrhizology is thriving as it contributes to both understanding of basic biology and to improving tree growing practice.

# References and notes.

(Many articles are accessible as abstract or full text using search engines such as Google Scholar, an institutional log-in, or a pay-wall.]

### Ecological context of roots and fungi.

Colinvaux, P., 1993. *Ecology 2*. John Wiley & Sons Inc., New York, ISBN: 0471558605. [Chapter 20, Ecological Succession, for discussion of ecosystem concept.]

Helm, D.J., Allen, E.B. & Trappe, J.M., 1999. Plant growth and ectomycorrhiza formation by transplants on deglaciated land near Exit Glacier, Alaska. Mycorrhiza, 8: 297-304.

Krebs, C.J., 2014. *Ecology: the experimental analysis of distribution and abundance.* Pearson Education Ltd. Harlow, U.K., ISBN: 9781292026275. [Pages 23-26 on scientific method as performed in ecology.]

Nara, K., 2006. Ectomycorrhizal networks and seedling establishment during early primary succession. New Phytologist, 169: 169-178.

Smith, S.E. & Read, D.J., 2008. *Mycorrhizal Symbiosis*. Academic Press, London. ISBN: 978-0-1237-0526-6. [Detailed and comprehensive textbook on all types of mycorrhizal symbiotic fungi.]

### Roots.

Baldrian, P., 2017. Forest microbiome: diversity, complexity and dynamics. FEMS Microbiology Reviews, 41: 109-130.

Chen, W., *et al.*, 2016. Root morphology and mycorrhizal symbioses together shape nutrient foraging strategies of temperate trees. Proceedings of the National Academy of Science of the USA, 113: 8741-8746.

Hirons, A.D. & Thomas, P.A., 2018. *Applied Tree Biology*. John Wiley & Sons Inc, Hoboken, USA. ISBN: 9781118296400. [Chapter 4 for general account of roots in context of silviculture and use of mycorrhizal inoculants.]

Thomas, P.A., 2014. *Trees: their natural history*. Cambridge University Press, Cambridge, UK. ISBN: 9780521133586. [Chapter 4 for general account of biology of roots.]

### Soil and fungi.

Anonymous, 2023. Articles: 'Soil', and 'Mycorrhiza' in Wikipedia. [Detailed introductions to these topics.]

Brundrett, M.C., 2002. Coevolution of roots and mycorrhizas of land plants. New Phytologist, 154: 275-304.

Högberg, M.N. & Högberg, P., 2002. Extramatrical ectomycorrhizal mycelium contributes one third of microbial biomass and produces, together with associated roots, half the dissolved organic carbon in a forest soil. New Phytologist, 154: 791-795.

Le Page, B.A., *et al.*, 1997. Fossil ectomycorrhizae from the Middle Eocene. American Journal of Botany, 84: 410-412.

Stubblefield, S.P., Taylor, T.N. & Trappe, J.M., 1987. Fossil mycorrhizae: a case for symbiosis. Science, 237: 59-60. [Detailed discussion of symbiosis as it occurs in arbuscular mycorrhorizas.]

Vogt, K.A., *et al.*, 1982. Mycorrhizal role in net primary production and nutrient recycling in *Abies amibilis* stands in western Washington. Ecology, 63: 370-380.

### Foraging for nutrients

Abuzinadah, R.A. & Read, D.J., 1989. The role of proteins in the nitrogen nutrition of ectomycorrhizal plants V. Nitrogen transfer in birch (*Betula pendula*) grown in association with mycorrhizal and non-mycorrhizal fungi. New Phytologist, 112: 61-68.

Chalot, M., *et al.*, 2002. An update on nutrient transport processes in ectomy-corrhizas. Plant and Soil, 244: 165-175.

Finlay, R.D. & Read, D.B., 1986. The structure and function of the vegetative mycelium of ectomycorrhizal plants. I. Translocation of <sup>14</sup>C labelled carbon between plants interconnected by a common mycelium. II. The uptake and distribution of phosphorus by mycelium interconnecting host plants. New Phytologist, 103:143-156; 157-165.

Nehls, U., 2008. Mastering ectomycorrhizal symbiosis: the impact of carbohydrates. Journal of Experimental Botany, 59: 1097–1108. [Well illustrated account of nutritional relationships of ectomycorrhizas.]

Quirk J., *et al.*, 2012. Evolution of trees and mycorrhizal fungi intensifies silicate mineral weathering. Biology Letters, 8: 1006-1011. [Information about how ectomycorrhizas leach out minerals from rocks and trade that resource with plants.]

Thorley, R.M.S., 2015. The role of forest trees and their mycorrhizal fungi in carbonate rock weathering and its significance for global carbon cycling. Plant, Cell and Environment, 38: 1947-1961. [Information on rock minerals going from soil to trees via ectomycorrhizas.]

### Mycorrhizas.

Downes, G.M., Alexander, I.J. & Cairney, J.W.G., 1992. Study of ageing of spruce *Picea sitchensis* ectomycorrhizas. I: morphological and cellular changes in mycorrhizas formed by *Tylospora fibrillosa* and *Paxillus involutus*. New Phytologist, 122:141-152. [Light, fluorescent, and electron microscopy of ectomy-corrhizas on spruce seedlings.]

Garcia, K., *et al.*, 2016. Take a trip through the plant and fungal transportome of mycorrhiza. Trends in Plant Science, 21: 937-950. [Review of molecular mechanisms of nutrient transport by mycorrhizas.]

Leake, J.R., *et al.*, 2004. Networks of power and influence: the role of mycorrhizal mycelium in controlling plant communities and agroecosystem functioning. Canadian Journal of Botany, 82: 1016-1045. [Detailed discussion of mycelial networks with some striking images of mycorrhizas.]

Massicotte, H.B., *et al.*, 1986. Structure and ontogeny of *Alnus crispa - Alpova diplophloeus* ectomycorrhizae. Canadian Journal of Botany, 64: 177-192.

Massicotte, H.B., Peterson, R.L. & Melville, L.H., 1989. Hartig net structure of ectomycorrhizae synthesized between *Laccaria bicolor* (Tricholomataceae) and two hosts: *Betula alleghaniensis* (Betulaceae) and *Pinus resinosa* (Pinaceae). American Journal of Botany, 76:1654-1667. [Many clear images of root-hairs and Hartig net in both these papers by Massicotte and co-authors.]

Plett, J. M. & Martin F., 2011. Blurred boundaries: lifestyle lessons from ectomycorrhizal fungal genomes. Trends in Genetics, 27: 15-22. [Account of recent findings on the genetic basis of mycorrhizal symbiosis.]

Söderström, B. & Read. D.J., 1987. Respiratory activity of intact and excised ectomycorrhizal mycelial systems growing in unsterilized soil. Soil Biology and Biochemistry, 19: 231-236.

### Symbiosis.

Anonymous, 2023. Articles: 'Coevolution', 'Mutualism', 'Holism in science'. Wikipedia.

Beiler, K.J., Simard, S.W. & Durall D.M., 2015. Topology of tree–mycorrhizal fungus interaction networks in xeric and mesic Douglas-fir forests. Journal of Ecology, 103: 616–628. [Use of network analysis to understand functioning of mycelial connections between trees.]

Bever, J.D., 2015., Preferential allocation, physio-evolutionary feedbacks, and the stability and environmental patterns of mutualism between plants and their root symbionts. New Phytologist, 205: 1503-1514.

Figueiredo, A.F., Boy, J. & Guggenberger, G., 2021. Common mycorrhizae network: a review of the theories and mechanisms behind underground interactions. Frontiers in Fungal Biology, 2: Article number 735299.

Frederickson, M. 2017 Mutualisms are not on the verge of breakdown. Trends in Ecology & Evolution, 32: 727-734. [Discussion of how mutualisms between fungi and plants can involve unequal sharing, otherwise known as cheating.]

Hoeksema, J.D. & Bruna, E.M., 2000. Pursuing the big questions about interspecific mutualism: a review of theoretical approaches. Oecologia, 125: 321-330. [Balance of increase in evolutionary fitness by trading resources.] Jones, M.D., Durall D.M. & Cairney, J.W.G., 2003. Ectomycorrhizal fungal communities in young forest stands regenerating after clearcut logging. New Phytologist, 157: 399-422. [Review of functioning of ectomycorrhizal symbioses in relation to management of forests.]

Jones, M.D. & Smith, S.E., 2004. Exploring functional definitions of mycorrhizas: Are mycorrhizas always mutualisms? Canadian Journal of Botany, 82: 1089-1109. [Discussion of mycorrhizal symbiosis as a continuum from parasitism to mutualism.]

Karst, J., Jones, M.D. & Hoeksema, J.D., 2023. Positive citation bias and overinterpreted results lead to misinformation on common mycorrhizal networks in forests. Nature Ecology and Evolution, 13 February, online, 11 pages. DOI: 10.1038/s41559-023-01986-1. [Critique of some aspects of interpretation of empirical results on how common mycelial networks relate to ecology of trees.]

Klein, T., Siegwolf, W. & Korner, C., 2016. Below ground carbon trade among tall trees in a temperate forest. Science, 352: 342-344. [Common mycelial networks connecting multiple trees forming a guild of symbionts.]

Leigh, E. G., 2010. The evolution of mutualism. Journal of Evolutionary Biology, 23: 2507-2528. [Review of mutualism in general, with direct relevance to mycorrhizal symbioses.]

Levin, S.A., 2003. Complex adaptive systems: exploring the known, the unknown and the unknowable. Bulletin (New Series) of the American Mathematical Society, 40: 3-19. [Ecosystem is an example of complex adaptive system, and insight into analytical methods used to understand such systems.]

Näsholm, T., *et al.*, 2013. Are ectomycorrhizal fungi alleviating or aggravating nitrogen limitation of tree growth in boreal forests? New Phytologist, 198: 214-221.

### Forestry and mycorrhizas.

Beiler, K.J., *et al.*, 2010. Architecture of the wood-wide web: *Rhizopogon* genets link multiple Douglas-fir cohorts. New Phytologist, 185: 543-553. [Field study of the distribution patterns of a common mycelial network.]

Brownlee, C., *et al.*, 1983. The structure and function of mycelial systems of ectomychorrizal roots with special reference to their role in forming inter-plant connections and providing pathways for assimilate and water transport. Plant and Soil, 71: 433-443.

Brundrett, M., *et al.*, 1996. *Working with mycorrhizas in forestry and agriculture. ACIAR Monograph 32*. Australian Centre for International Agricultural Research, Canberra. ISBN: 1863201815. [Instruction manual for studying mycorrhizas, and their applications to forestry.]

Dey, D.C. & Parker, W.C., 1996. Regeneration of red oak (*Quercus rubra*) using shelterwood systems, ecophysiology, silviculture and management recommendations. Forest Research Information Paper No. 126. Ontario Ministry of Natural Resources, Canada.

Jones, M.D., 2017. Integrating ectomycorrhizas into sustainable management of temperate forests. Chapter 11, pgs 187-211, in: *Mycorrhizal Mediation of Soil: fertility, structure and carbon storage*. Johnson, N.C. *et al.* (eds.). Elsevier, Amsterdam. ISBN: 9780128043127.

Marx, D.H., Marrs, L.F. & Cordell, C.E., 2002. Practical use of mycorrhizal fungal technology in forestry, reclamation, aboriculture, agriculture and horticulture. Dendrobiology, 47: 29-42.

Menkis, A., *et al.*, 2005. Fungal communities in mycorrhizal roots of conifer seedlings in forest nurseries under different cultivation systems, assessed by morphotyping, direct sequencing and mycelial isolation. Mycorrhiza, 16: 33-41.

Simard, S.W., 2009. The foundational role of mycorrhizal networks in selforganization of interior Douglas-fir forests. Forest Ecology and Management, 258S: S95-107. [Extensive review of knowledge on mycorrhizas in forests, plus applications to forestry.]